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Diazinon Properties, Mode of Action, and Toxicological Effects on Non-target Organisms

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Abstract

Diazinon (DZN) (O, O-diethyl-O-[2-isopropyl-6-methyl-4-pyrimidinyl] phosphorothioate) is one of the most widely applied organophosphorus pesticides in the livestock industry. It is an anticholinesterase organophosphate insecticide that inhibits the action of acetylcholinesterase, resulting in an abundance of acetylcholine, which affects neuromuscular transmission and causes insect paralysis and death. However, DZN residues in the environment can affect non-target organisms through the air, water, soil, and food chain, which raised worries about its detrimental consequences. Oxidative stress (OS) caused by reactive

oxygen species may be the key mechanism that might be related to DZN toxicity. This article provides a comprehensive investigation of DZN's toxicity pattern, with an emphasis on its negative effects on the neurological, hepatic, cardiac, and nephrotic systems, as well as its effects on reproductive processes. It also addresses how DZN affects hormonal levels, causing endocrine disorders. Consequently, identifying these pathways might offer helpful insight on the mechanisms of DZN activity on various tissues. This review aimed to raise awareness about the harmful effects of DZN exposure.

Keywords: Diazinon, Contaminant, AchE, Oxidative Stress, Organs Toxicity

Introduction

Pesticides are a class of chemicals used to eradicate insects, weeds, fungus, and bacteria ^[1-2]. The use of pesticides in veterinary field is economically required. There are essential to control insects and parasites that impair livestock production, consume crops used for animal feed, and spread illnesses ^[3]. The vast majority of pesticides used in the veterinary medicine are ectoparasiticides, which target insects that live on animals' skin. Ectoparasiticide active components are commonly applied to animals' skins in a range of formulations ^[4].

Organophosphates (OPs) pesticides are one of the most frequently used pesticides. They are esters, amides, or thiols derived from phosphoric, phosphonic, phosphinic, or thiophosphoric acids, coupled with two organic groups and a side chain containing cyanide, thiocyanate, or phenoxy groups ^[5-6]. These pesticides suppress the function of acetylcholinesterase, an enzyme that degrades acetylcholine, a neurotransmitter that controls the nervous system. Organophosphates block acetylcholinesterase, causing acetylcholine buildup that can lead to nervous system overstimulation and, finally, paralysis and death of the target insect ^[7]. Even with its benefits, its extensive use has killed non-target creatures, contaminated the environment, and caused acute and severe chronic toxicity in many different parts of the world. Thus, it has been considered a health risk to the environment ^[8].

Among the most popular organophosphate pesticides is diazinon (DZN). It is a popular anti-parasitic medication used to combat external parasites like ticks and mites ^[9]. The present review will focus on current information about DZN, its physical and chemical properties, and the broad toxicological impacts of DZN, providing insight into the processes behind these effects.

a. Chemical Structure

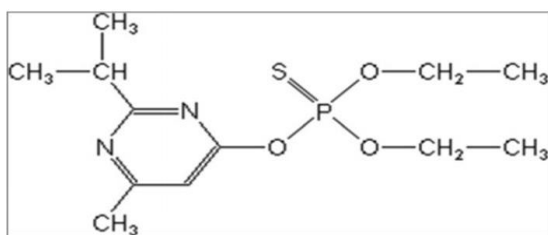


Fig 1: Chemical structure of diazinon^[10]

The organophosphorus insecticide diazinon (Fig 1) is a colourless to dark brown liquid with a density of 1.116 g/cm³, which is more than that of water. DZN has the Chemical Abstracts Service (CAS) registration number 333-41-5. The CAS name for diazinon is O, O-diethyl O-[6-methyl-2-(1-methylethyl)-4-pyrimidinyl] phosphorothioate.

b. DZN Uses

Diazinon is often used in veterinary medicine as a nematicide, acaricide, and insect repellent sprayed on poultry and animals^[11-12]. DZN comes in a variety of forms, including dusts, granules, emulsions, oily solutions, microcapsules, ribbons, tags, aerosols, and seed dressings^[13]. There are currently more than 500 diazinon-containing products available on the market, either alone or in combination with other pesticides^[14].

c. Diazinon properties

The water solubility of DZN is rather high at 60 mg/L, and its boiling point is between 82 and 84 °C^[14]. It takes 138 days to fully hydrolyse, whereas its half-life in soil was around 40 days. It is stable at pH 7 and does not easily volatilise from water and soil. Therefore, it can remain in the environment for up to six months^[15]. DZN is oxidisable and decomposes at temperatures over 120 °C^[16]. However, DZN degradation byproducts are harmful to both human and environmental health^[17]. Furthermore, according to Begum *et al.*^[18], DZN is classified as a nonsystemic (contact or surface) pesticide in nature.

DZN is one of the most frequently found OPs in groundwater, drinking water, and surface water, which is a particularly serious issue^[19], as Pesticides with high water solubility may diffuse under the root area, sometimes contaminating groundwater, or get spread by surface drainage far from their use location^[20].

d. DZN mode of action

Diazinon can affect a pest's body through contact, ingestion, or respiratory pathway^[21]. The toxicity mechanism of DZN on insects is similar to that of other organophosphorus insecticides. Its mode of action relies on the phosphorylation of the amino acid serine in the active site of the acetylcholinesterase enzyme, which inhibits the cholinesterase enzyme^[22]. AChE inhibition causes acetylcholine to build up at synapses between neurones, which prolongs the cholinergic receptors stimulation. Consequently, the continuous neurone stimulation causes intermediate symptoms such as anorexia, diarrhoea, generalised weakness, muscular tremors, altered posture and behaviour, depression, and insect mortality^[23].

Despite the benefits of diazinon, it may be hazardous to non-target species^[24]. Several investigations have shown

residues of DZN in soil, water, and foods such as fruits, vegetables, and milk^[25]. DZN converted into more hazardous degradate diazoxon by oxidative desulfurization, an activation process, after it has been ingested by both target and non-target species. Animals exposed to both the parent residue and the very poisonous oxon form as a result of the parent compound's metabolism to the oxon^[22]. Diazoxon not only inhibits cholinesterase but also increases reactive oxygen species (ROS), leading to oxidative stress. Oxidative stress releases DNA and ruptures the mitochondrial membrane, causing cytotoxicity in different tissues^[26].

e. DZN metabolism and toxicity

DZN is now being utilised in the Middle East in the veterinary and agricultural sectors, despite being prohibited in some countries^[27]. The World Health Organisation classified it as a class II pollutant that is moderately dangerous^[27-28]. It is important to emphasise that DZN has a closer link with insect AChE than with mammalian homologue enzymes. This fact results in significant variances in the LD50 dosage of DZN for mammals and insects^[13]. Diazinon's toxicity to all animal species is dose-dependent; the larger the dose, the higher the mortality rate, coupled by a shorter mortality period^[29].

DZN poisoning in animals happens as a result of contaminated food and water. After oral intake, DZN is quickly absorbed and dispersed throughout the body and undergoes extensive metabolism^[30]. Cytochrome enzymes, also known as monooxygenase enzymes, such as CYP2C19, CYP1A2, CYP2B6, and CYP3A4, use oxidative desulfuration to convert DZN to diazoxon. These reactions occur mostly in the cells of the liver and intestines within the microsomal endoplasmic reticulum. Liver microsomes have a 5- to 10-fold greater DZN metabolism than enterocyte microsomes^[13-14].

Liver is the primary organ involved in CYP metabolism and is in charge of the metabolism of pesticides that are ingested from the gastrointestinal system^[31]. DZN and diazoxon may also be detoxified (deactivated) by microsomal enzymes like CYPs and esterases to produce the deactivated and non-toxic compounds 2-isopropyl-4-methyl-6-hydroxypyrimidine (IMHP), diethyl phosphate (DEP), and diethyl thiophosphate (DETP). Afterward, they were mostly eliminated in urine^[30]. Accordingly, the toxicity resulting from diazinon exposures to non-target organisms will be determined by the equilibrium between activation and detoxification processes^[32].

Since DZN is one of OPs, it exhibits the same general symptoms of OPs poisoning, which include emesis, stomach cramps, lacrimation, urine, faeces, and salivation. These symptoms are referred to as "SLUDGE" symptoms^[33-34]. Furthermore, DZN's lipophilicity makes it easier for it to interact with the phospholipid bilayer of cell membranes of the majority of visceral organs^[35]. As a result, DZN is harmful to several organs through the production of oxidative stress. According to several studies, it is a compound that is known to be cardiotoxic, immunotoxic, genotoxic, neurotoxic, hepatotoxic, cytotoxic, nephrotoxic, genotoxic, cytotoxic, cytotoxic, nephrotoxic, renal, and reproductive toxicity^[36-42].

e.1 DZN-induced oxidative stress

A large number of diazinon's deleterious effects are produced by oxidative stress in addition to the suppression

of the acetylcholinesterase enzyme [43]. *In vitro* and *in vivo* studies have validated the significance of oxidative stress in the pathophysiology of DZN-induced tissue injuries [42]. Through the generation of reactive oxygen species, which in turn causes oxidative stress on glycolysis, mitochondrial respiratory tract, ATP production, amino acid metabolism, antioxidant defence system, and poison detoxification of the cell [44-45]. The incidence of oxidative stress following various DZN administration protocols has been examined in a number of studies. These protocols may include a single administration of increasing doses or after their prolonged administration (subacute, subchronic, or chronic) [46-49].

Previous findings reported that acute or subacute DZN poisoning causes lipid peroxidation (LPO) of cell membranes in the rat brain, heart, spleen, liver, and kidneys. This is characterised by a notable alteration in the activity of the antioxidant enzymes superoxide dismutase (SOD), catalase (CAT), and glutathione-S-transferase (GST), followed by alterations in lactate dehydrogenase (LDH) activity and malondialdehyde (MDA), and glutathione (GSH) levels [24, 25, 50, 51, 52]. Moreover, DZN causes oxidative damage to proteins with increases in the concentration of protein carbonyl groups and also increases Nitrite (NO₂) concentrations [53-56].

DZN has been demonstrated to affect DNA integrity and induce cellular dysfunction by producing DNA adducts and lesions, single-strand and double-strand DNA breaks, and DNA and protein inter- and intra-crosslinks [57]. Furthermore, DZN exposure rises the frequency of micronuclei (genetic damage) produced by DNA strand breaks and oxidative stress-induced chromosomal abnormalities [58]. Studies by Naderi *et al.* and Ali *et al.* [59, 60] found that DZN intoxication promoted DNA fragmentation testicular spermatogenic tissue and hepatic tissues in mice. This degradation may be attributed to alkylating properties of organophosphorus chemicals.

e.2 DZN hepatotoxicity

The liver is essentially the first organ to experience severe oxidative stress from diazoxon because liver microsomal CYP450 enzymes convert DZN to diazoxon through oxidative desulphurization [61]. Given the liver's primary function in DZN metabolism and the correlation between DZN and oxidative stress, it can be concluded that oxidative stress plays a key role in DZN-induced liver damage [54].

Lipid peroxidation causes cell membrane disintegration, membrane fluidity reduction, membrane permeability enhancement, and cytoplasmic enzyme leakage [62]. The antioxidant system's depletion therefore causes hepatocellular death and necrosis, which is followed by a rise in the intracellular hepatic enzymes (ALT, AST, ALP, γ -GT, and LDH activities). The measurement of antioxidant levels for the evaluation of hepatocellular damage verified these increases. Additionally, histological techniques can reveal the extent of liver damage [63-64]. It was demonstrated that after diazinon administration, rats' livers exhibit periportal inflammation and necroinflammatory alterations [65]. Numerous publications have also shown hepatotoxicity and pathohistological alterations following acute [66], subacute [67-69], and subchronic exposure to diazinon in rats [70].

Previous research by Nassar *et al.* [58] found that oral treatment of DZN greatly elevated Bax, Caspase-3, Caspase-9, and the Bax/Bcl-2 ratio, whereas Bcl-2 dropped dramatically in liver and also protein carbonyl (PC) risen

significantly followed by substantial declines in SOD, CAT, GST, and GPx levels. Fat change and hepatocyte necrosis can occur as a result of DZN's capability to impede protein synthesis and hydrolysis [71].

e.3 DZN nephrotoxicity

The kidney is a vital organ that performs a number of vital tasks, such as preserving homeostasis, controlling the extracellular environment, detoxifying the body, and getting rid of harmful substances and drugs. As a result, toxins may target the kidney [72]. DZN and its metabolites may be linked to kidney and renal tubule damage [73]. Rats given DZN showed large increases in MDA, H₂O₂, and NO in their renal tissue, which are indicators of oxidative and nitrosative stress, which is the imbalance between the production of reactive oxygen or nitrogen species and the antioxidant defence system. This imbalance promotes the generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS) over the antioxidant defence system [74]. The RNS and ROS produced by DZN may impair the kidney's capacity to eliminate sodium, which results in sodium retention and hypertension. A decrease in the glomerular filtration rate (GFR) and an increase in salt retention impair the nephron's functional ability [74]. Additionally, as blood creatinine and urea nitrogen levels roughly correspond to the glomerular filtration rate, a rise in these parameters following DZN treatment indicated a severe deterioration in renal function [75-76].

Renal histological alterations, including glomerular degeneration and kidney parenchymal shrinkage with leukocytic infiltration and congestion, were seen in many studies in experimental animals exposed to DZN. In addition, renal tubules had degenerative and necrotic alterations, including eosinophilic cytoplasm, broken cell borders, and blood capillary congestion between degenerated tubules [37, 67, 76]. Furthermore, histological analyses showed that the cortex was more damaged than the medulla. This may be partially caused by the unequal distribution of DZN and its metabolites in renal tissue, where the circulation carries approximately 90% of the entire renal blood flow to the cortex. Thus, the cortex may receive a higher bloodstream concentration of DZN and its metabolites than the medulla [74].

e.4 DZN reproductive toxicity

One of the primary targets of DZN toxicity is the reproductive system. DZN causes degenerative alterations in testicular tissue [77], a significant decrease in Leydig, spermatogonia, primary spermatocytes, and spermatid cells, poor sperm quality [78], and a decrease in serum testosterone levels that results in secondary infertility [79, 43]. Key cause of DZN reproductive toxicity is oxidative stress (OS) [80]. The DZN effect in the testis may target developing spermatozoa [81]. Through a variety of processes, it causes chromosomal damage, abnormalities, or the breakdown of sperm proteins, which eventually lowers sperm parameters [80]. DZN induced Sperm DNA fragmentation was previously reported by Salazar-Arredondo *et al.* [82] and Piña-Guzman *et al.* [81] as DZN exposure leads to nuclear protamine phosphorylation, which alters sperm chromatin condensation and DNA integrity.

Numerous experimental investigations have demonstrated that DZN can cause mitochondrial damage in sperm by reducing mitochondrial activity and causing the loss of mitochondrial membrane potential, affecting sperm motility, capacitation, and fertility rate [83-84]. This can be explained as

the proper function of sperm and the fertilisation process depend on mitochondria. They produce ROS and ATP, which are essential for healthy sperm activity and capacitation^[85]. Additionally, they act as intracellular Ca²⁺ stores, and the membrane potential of these cells is essential for maintaining sperm motility and energy^[86]. Histological study shows that animals intoxicated with DZN have minimal cytoplasmic carbohydrate accumulation in their spermatogenic cells. The shortage of energy sources may have caused the spermatogenesis cells to switch from using glucose to lipids^[87].

Research conducted by Abou Hasan *et al.*^[88] on rats to assess the toxicity of DZN on female rats' ovaries reported that a single dosage of DZN enhanced the amount of cells undergoing apoptosis and accelerated the atresia process in atretic follicles. Although the corpus luteum cells of the DZN-intoxicated rats showed multilamellar body forms and accumulations of big lipid droplets. DZN inhibited steroidogenesis in the rat ovary and STAR mRNA expression in cells affecting female fertility^[89], as the transfer of lipids from the outer mitochondrial membrane to the inner membrane, a crucial step in the synthesis of steroid hormones, is regulated by the STAR protein, which also controls the luteinisation process of granulosa lutein and theca lutein cells^[90].

e.5 DZN neurotoxicity

DZN was the first organophosphate to be recognised as a developmental neurotoxicant^[91]. Irreversible cholinesterase inhibition is the main target of DZN and its metabolites. DZN's suppression of AChE then causes ACh to build up at the connections between neurones, which raises the excitability of cholinergic receptors^[92]. The neurotransmitter systems balance in cortical and subcortical areas is simply disrupted by excess ACh, which also modifies the ACh feedback loop^[93-94]. According to Ahmed *et al.*^[95], imbalances between provocative and suppressive neurones in various brain regions are another theory that might account for neurological consequences. Ultimately leading to muscarinic and nicotinic symptoms as well as indicators of intoxication in the central and peripheral nervous systems such as general weakness, anorexia, muscle convulsions, diarrhoea, abnormal behaviour, depression, and mortality after extended DZN exposure^[14, 96]. Diazoxon is a more powerful AChE inhibitor in nerve tissues than its parent chemical, DZN (more than 1000 times)^[96]. Prenatal DZN exposure can result in significant alterations in brain function. Studies identified deficits in glutamatergic^[97], serotonergic, and cholinergic functions^[98] in the brains of exposed kids. These neurochemical alterations are linked to numerous behavioural deficiencies, such as teenage hyperactivity and decreased risk-avoidance^[99], poor passive avoidance learning^[100], and changes in novel object recognition^[97-99].

DZN exposure was reported to change expression of the hippocampus genes important for neurotransmission. It causes downregulation of the genes producing GABAB1 receptors, which are a key contender in the establishment of recognition memory^[101]. Moreover, DZN exposure caused down regulation of the synaptophysin (SYP) gene, resulting in synaptic degeneration and substantial synapse loss in mice^[102]. Also, Afshari *et al.*^[103] found elevated TNF- α levels in the rat prefrontal cortex after 5 days of DZN administration.

e.6 DZN cardiotoxicity

Numerous investigations have revealed that cardiotoxicity is linked to DZN toxicity^[104-105]. DZN caused left ventricular hypertrophy and decreased myocardial contractility, which led to heart failure^[74]. It has been proposed that lipid peroxidation is one of the molecular processes behind DZN-induced cardiac damage^[105]. Increases in the aminotransferases ALT and especially AST as well as alkaline phosphatase are indicators of cardiac injury, indicating that DZN damaged the pericardial membranes, allowing these enzymes to increase in the blood^[74]. Additionally, it suggested that DZN may promote the generation of O₂^{•-} by NADPH oxidase^[106]. O₂^{•-} and NO react to generate ONOO⁻, which lowers NO's bioavailability and eventually results in hypertension. According to Ajibade *et al.*^[74], NO is known to control cardiac contractility, restrict leukocyte adherence to the endothelium, influence vascular tone, and inhibit platelet activation.

e.7 DZN as endocrine disruptor

DZN is classified as an endocrine disruptor "having the potential to cause endocrine disruption."^[77] Pesticides with endocrine-disrupting activity alter an animal's endocrine system by interfering with receptor binding and disrupting hormone metabolism and steroidogenesis^[107]. They have oestrogenic and antiandrogenic properties that can change the sex-steroid synthesis enzymes by acting on the hypothalamo hypophyseal-gonadal axis (HHG)^[108]. This hormonal change modulates development and gonadal growth, which can lead to ovarian or testicular dysfunction^[109]. DZN has been shown in recent studies to disrupt sex hormone levels, including testosterone and gonadotropin follicle-stimulating hormone (FSH), and luteinizing hormone (LH), which are the primary regulators of spermatogenesis and germ cell development^[110].

e.8 DZN induced apoptosis and inflammation

Numerous cell types, such as ovarian follicular cells, sperm cells, cardiac muscle cells, peripheral blood lymphocytes, and skeletal muscles, have been shown to undergo DZN-induced cell death and apoptosis^[83, 86, 111]. One of the primary ways that DZN causes apoptosis is thought to be through increased activity of caspases-3, -8, and -9, as well as increased Bax contents and decreased Bcl-2 (improved Bax/Bcl-2 ratio)^[10]. Another important mechanism by which DZN promotes apoptosis is oxidative stress brought on by ROS^[112]. Through the release of mitochondrial cytochrome c and the stimulation of caspases, the main initiators of apoptosis, ROS cause cell damage and death^[113].

Pro-inflammatory cytokines such as IL-1 β , IL-10, and TNF- α have been shown to rise in response to DZN^[114-116]. This might be because DZN directly raises the mRNA expression of pro-inflammatory cytokines^[117] or because the oxidative stress caused by DZN exposure triggered an inflammatory response^[118].

e.9 DZN immunotoxicity

Exposure to diazinon has immunotoxic effects that are likely caused by changes in immune-related gene expression and modulation of the main cytokines^[119]. Mice exposed to DZN have been shown to have histopathological alterations in the thymus and spleen as well as inhibition of humoral and cellular immune system function^[120]. Additionally, phagocytic indices and splenocyte proliferation were significantly reduced in Nile tilapia exposed to DZN,

indicating immunotoxicity on both the innate and acquired immune responses^[121-122].

In the great sturgeon fish, sub-lethal levels of DZN had an impact on innate immunological markers such as lysozyme and white blood cells. Additionally, Persian sturgeon fish exposed to sub-lethal dosages of DZN showed a reduction in lymphocytes and an increase in total neutrophils^[123].

e.10 DZN induced hyperglycaemia

Prior research has documented hyperglycaemic and haemostatic alterations associated with glucose metabolism in DZN-exposed persons^[40, 54]. The hyperglycemia impact of DZN is caused by two primary pathways. The first is the accelerated rate of gluconeogenesis, which produces glucose from non-carbohydrate sources by influencing glucose-6-phosphatase [G6Pase] and phosphoenolpyruvate carboxykinase [PEPCK]. Second, DZN-induced insulin resistance results from the decrease of insulin receptor expression and glucose transporter type 4 (GLUT4) because adipocytes secrete pro-inflammatory adipokines like TNF- α . This, in turn, causes a decrease in glucose uptake in adipose tissue^[124].

AChE inhibition, a primary mechanism of DZN toxicity, can also increase the synthesis of adrenocorticotrophic hormone (ACTH), which controls the production of cortisol. This hormone promotes gluconeogenesis and disrupts the action of insulin^[125-127]. Animals exposed to DZN experience dose-dependent hyperglycemia. Blood glucose levels may also be impacted by an animal's gender; females are more likely to experience hyperglycemia after being exposed to DZN^[128].

Conclusion

Diazinon (DZN) is an organophosphate insecticide frequently utilized to control insects, ticks, and mites. Its exposure may lead to OP toxicity in non-target organisms. OS induced by reactive oxygen species is the main mechanism associated with DZN toxicity. DZN has multi organ toxicity such as hematological and reproductive disorders, as well as kidney, liver, immuno-toxicity, and central nervous system toxicity. Understanding the harmful effects of DZN is critical for reducing their influence on animal health and directing future research efforts.

List of abbreviations

ACh	Acetyl Choline
AChE	Acetyl Choline Esterase
ACTH	Adrenocorticotrophic hormone
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
ATP	Adenosine triphosphate
Bax	BCL2 Associated X, Apoptosis Regulator
BCL2	Apoptosis Regulator
CAS	Chemical Abstracts Service
CAT	Catalase
CYP	Cytochromes P450
DEP	Diethyl phosphate
DETP	Diethyl thiophosphate
DZN	Diazinon
FSH	Follicle-stimulating hormone
G6Pase	Glucose-6-phosphatase
GFR	Glomerular filtration rate
GLUT4	Glucose transporter type 4
GPx	Glutathione peroxidase

GSH	Glutathione
GST	Glutathione s-transferase
H ₂ O ₂	Hydrogen peroxide
HHG	Hypothalamo hypophyseal-gonadal axis
IL-10	Interleukin 10
IL-1 β	Interleukin-1 beta
IMHP	2-isopropyl-4-methyl-6-hydroxypyrimidine
LDH	Lactate dehydrogenase
LH	Luteinizing hormone
LPO	Lipid peroxidation
MDA	Malondialdehyde
mRNA	Messenger ribonucleic acid
NADPH	Nicotinamide adenine dinucleotide phosphate
Nitrite	NO ₂
No	Nitrogen monoxide
O ₂ -	Superoxide radical
ONOO-	Peroxynitrite
OPs	Organophosphate pesticides
OS	Oxidative stress
PC	protein carbonyl
PEPCK	Phosphoenolpyruvate carboxykinase
RNS	Reactive nitrogen species
ROS	Reactive oxygen species
SOD	Superoxide dismutase
STAR	Steroidogenic Acute Regulatory Protein
TNF- α	Tumor necrosis factor-alpha
γ -GT	Gamma-glutamyltransferase

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Conflict of Interest

The authors declare that they have no conflict of interest.

References

- Jayaraj R, Megha P, Sreedev P. Organochlorine pesticides, their toxic effects on living organisms and their fate in the environment. *Interdisciplinary Toxicology*. 2016; 9(3-4):90-100.
- Georgiadis N, Tsarouhas K, Tsitsimpikou C, Vardavas A, Rezaee R, Germanakis I, *et al.* Pesticides and cardiotoxicity. Where do we stand. *Toxicology and Applied Pharmacology*. 2018; 353:1-14.
- Abubakar Y, Tijjani H, Egbuna C, Adetunji CO, Kala S, Kryeziu TL, *et al.* Pesticides, history, and classification. In *Natural remedies for pest, disease and weed control*. Academic Press, 2020, 29-42.
- Copland A, Elsheikha H. A snapshot of the adverse effects of companion animal ectoparasiticide. *Companion Animal*. 2021; 26(7):153-160.
- Balali-Mood M, Abdollahi M. Basic and clinical toxicology of organophosphorus compounds. London, Springer International Publishing, 2014.
- Kaushal J, Khatri M, Arya SK. A treatise on Organophosphate pesticide pollution: Current strategies and advancements in their environmental degradation and elimination. *Ecotoxicology and Environmental Safety*. 2021; 207:111483.
- Könemann S, von Wyl M, Vom Berg C. Zebrafish larvae rapidly recover from locomotor effects and neuromuscular alterations induced by cholinergic insecticides. *Environmental Science and Technology*. 2022; 56(12):8449-8462.
- Al-Rawashdeh M, Keikhosrokiani P, Belaton B, Alawida M, Zwiri A. IoT adoption and application for

- smart healthcare: A systematic review. *Sensors*. 2022; 22(14):5377.
9. Oñate E, Rezola E, Hernandez U, Muñoz JA. Organophosphate poisoning after using diazinon as an antiparasitic. In *Anales de Pediatría*. 2009; 71(3):272-273.
 10. Boussabbeh M, Ben Salem I, Hamdi M, Ben Fradj S, Abid-Essefi S, Bacha H. Diazinon, an organophosphate pesticide, induces oxidative stress and genotoxicity in cells deriving from large intestine. *Environmental Science and Pollution Research*. 2016; 23(3):2882-2889.
 11. Dorraki N, Mahdavi V, Ghomi H, Ghasempour A. Elimination of diazinon insecticide from cucumber surface by atmospheric pressure air-dielectric barrier discharge plasma. *Biointerphases*. 2016; 11(4):041007.
 12. Rahimnejad M, Abdulkareem RA, Najafpour G. Determination of Diazinon in fruit samples using electrochemical sensor based on carbon nanotubes modified carbon paste electrode. *Biocatalysis and Agricultural Biotechnology*. 2019; 20:101245.
 13. Debski B, Kania BF, Kuryl T. Transformations of diazinon, an organophosphate compound in the environment and poisoning by this compound. *ECOLOGICAL-BRATISLAVA*. 2007; 26(1):68-82.
 14. Aggarwal V, Deng X, Tuli A, Goh KS. Diazinon-chemistry and environmental fate: A California perspective. *Reviews of Environmental Contamination and Toxicology*. 2013; 223:107-140.
 15. DiBartolomeis M, Kegley S, Mineau P, Radford R, Klein K. An assessment of acute insecticide toxicity loading (AITL) of chemical pesticides used on agricultural land in the United States. *PLoS ONE*. 2019; 14(8):e0220029.
 16. Shayeghi M, Dehghani M, Mahvi A, Azam K. Application of acoustical processor reactors for degradation of diazinon from surface water. *Iran J Arthropod Borne Dis*. 2010; 4(2):11-18.
 17. Wang CK, Shih YH. Facilitated ultrasonic irradiation in the degradation of diazinon insecticide. *Sustainable Environment Research*. 2016; 26(3):110-116.
 18. Begum A, Ahmed MS, Alam SN. Decontamination methods for reduction of insecticide residues in Brinjal and Chilli. *International Journal of Agronomy and Agricultural Research*. 2016; 9(4):24-30.
 19. Glinki DA, Purucker ST, Van Meter RJ, Black MC, Henderson WM. Analysis of pesticides in surface water, stemflow, and throughfall in an agricultural area in South Georgia, USA. *Chemosphere*. 2018; 209:496-507.
 20. De Souza RM, Seibert D, Quesada HB, de Jesus Bassetti F, Fagundes-Klen MR, Bergamasco R. Occurrence, impacts and general aspects of pesticides in surface water: A review. *Process Safety and Environmental Protection*. 2020; 135:22-37.
 21. Nematollahi A, Rezaei F, Afsharian Z, Mollakhalili-Meybodi N. Diazinon reduction in food products: A comprehensive review of conventional and emerging processing methods. *Environmental Science and Pollution Research*. 2022; 29(27):40342-40357.
 22. Wu X, Li J, Zhou Z, *et al*. Environmental occurrence, toxicity concerns, and degradation of diazinon using a microbial system. *Frontiers in Microbiology*. 2021; 12.
 23. Larkin DJ, Tjeerdema RS. Fate and effects of diazinon. *Reviews of environmental contamination and toxicology*. 2000; 166:49-82.
 24. Jafari M, Salehi M, Ahmadi S, Asgari A, Abasnezhad M, Hajigholamali M. The role of oxidative stress in diazinon-induced tissues toxicity in Wistar and Norway rats. *Toxicology Mechanisms and Methods*. 2012; 22(8):638-647.
 25. Tari K, Samarghandi MR, Fard NJH, Jorfi S, Yari AR, Fard MP. Pollution Status of Pesticide Residues in Food Products in Iran: A Mini-review within 2008-2018. *Archives of Hygiene Sciences*. 2020; 9(3):214-223.
 26. Hosseini-Rahbari A, Nazem H, Fazilati M, Mehri F. Protective effect of resveratrol against sub-acute diazinon-induced oxidative stress in rat kidney. *Koomesh*. 2021; 23(6):794-800.
 27. Pirsahab M, Dargahi A, Hazrati S, Fazlzadehdavil M. Removal of diazinon and 2,4-dichlorophenoxyacetic acid (2,4-D) from aqueous solutions by granular-activated carbon. *Desalination and Water Treatment*. 2013; 52(22-24):4350-4355.
 28. Jonidi-Jafari A, Shirzad-Siboni M, Yang JK, Naimi-Joubani M, Farrokhi M. Photocatalytic degradation of diazinon with illuminated ZnO-TiO₂ composite. *Journal of the Taiwan Institute of Chemical Engineers*. 2015; 50:100-107.
 29. Solymann MSM, Boshahma FH, Mohamed H, Elmhalli FH, Amer AH. Assessment of Lethal Dose and Lethal Time of Diazinon in Swiss Albino Mice. *Journal of Clinical Epidemiology and Toxicology*. 2024; 5(4):2-5.
 30. Burgess P, Harper C, Todd GD, Wohlers D. Toxicological profile for diazinon, 2008.
 31. Poet TS. *In vitro* rat hepatic and intestinal metabolism of the organophosphate pesticides chlorpyrifos and diazinon. *Toxicological Sciences*. 2003; 72(2):193-200.
 32. Ellison CA, Tian Y, Knaak JB, Kostyniak PJ, Olson JR. Human hepatic cytochrome P450-Specific metabolism of the organophosphorus pesticides methyl parathion and diazinon. *Drug Metabolism and Disposition*. 2011; 40(1):1-5.
 33. Rastogi S, Tripathi S, Ravishanker D. A study of neurologic symptoms on exposure to organophosphate pesticides in the children of agricultural workers. *Indian Journal of Occupational and Environmental Medicine*. 2010; 14(2):54.
 34. Peter JV, Sudarsan T, Moran J. Clinical features of organophosphate poisoning: A review of different classification systems and approaches. *Indian Journal of Critical Care Medicine*. 2014; 18(11):735-745.
 35. Videira RA, Antunes-Madeira MC, Lopes VI, Madeira VM. Changes induced by malathion, methylparathion and parathion on membrane lipid physicochemical properties correlate with their toxicity. *Biochim Biophys Acta*. 2001; 1511(2):360-368.
 36. Razavi B, Hosseinzadeh H, Imenshahidi M, Malekian M, Ramezani M, Abnous K. Evaluation of protein ubiquitylation in heart tissue of rats exposed to diazinon (an organophosphate insecticide) and crocin (an active saffron ingredient): Role of HIF-1A. *Drug Research*. 2014; 65(11):561-566.
 37. Mansour SA, Abbassy MA, Shaldam HA. Hepato-renal toxicity induced by chlorpyrifos, diazinon and their mixture to male rats with special concern to the effect

- of zinc supplementation. *J Toxicol Pharmacol.* 2017; 1(3):15-24.
38. Karimani A, Heidarpour M, Jafari AM. Protective effects of glycyrrhizin on sub-chronic diazinon-induced biochemical, hematological alterations and oxidative stress indices in male Wistar rats. *Drug and Chemical Toxicology.* 2018; 42(3):300-308.
 39. Nili-Ahmadabadi A, Ali-Heidar F, Ranjbar A, *et al.* Protective effect of amlodipine on diazinon-induced changes on oxidative/antioxidant balance in rat hippocampus. *Research in Pharmaceutical Sciences.* 2018; 13(4):368.
 40. Nili-Ahmadabadi A, Akbari Z, Ahmadimoghaddam D, Larki-Harchegani A. The role of ghrelin and tumor necrosis factor alpha in diazinon-induced dyslipidemia: insights into energy balance regulation. *Pesticide Biochemistry and Physiology.* 2019; 157:138-142.
 41. Anbarkeh FR, Nikravesht MR, Jalali M, Sadeghnia HR, Sargazi Z. The effect of diazinon on cell proliferation and apoptosis in testicular tissue of rats and the protective effect of vitamin E. *PubMed.* 2019; 13(2):154-160.
 42. Esfahani M, Mehri F. Homeostatic changes of trace elements in diazinon toxicity in rat model: The beneficial role of resveratrol. *Toxicology Reports.* 2024; 13:101719.
 43. Salahi S, Ghazanfari R, Darwish A, Mazlounshahraki R, Shakibzade Y, Kord H, *et al.* The Effect of Diazinon Poison on Changes in Testicular Tissue and Testosterone Hormone levels in NMRI Lab Mice. *Journal of Biochemical Technology,* 2018, 29-22.
 44. Allen JW, Wolf DC, George MH, *et al.* Toxicity Profiles in Mice Treated with Hepatotumorigenic and Non-Hepatotumorigenic Triazole Conazole Fungicides: Propiconazole, Triadimefon, and Myclobutanil. *Toxicologic Pathology.* 2006; 34(7):853-862.
 45. Taxvig C, Hadrup N, Boberg J, *et al.* *In vitro* - *in vivo* correlations for endocrine activity of a mixture of currently used pesticides. *Toxicology and Applied Pharmacology.* 2013; 272(3):757-766.
 46. Fukai T, Ushio-Fukai M. Superoxide dismutases: Role in redox signaling, vascular function, and diseases. *Antioxidants and redox signaling.* 2011; 15(6):1583-1606.
 47. Kodydková J, Vávrová L, Kocík M, Zak A. Human catalase, its polymorphisms, regulation and changes of its activity in different diseases. *Folia biologica.* 2014; 60(4):153-167.
 48. Porokhovnik LN, Passekov VP, Gorbachevskaya NL, Sorokin AB, Veiko NN, Lyapunova NA. Active ribosomal genes, translational homeostasis and oxidative stress in the pathogenesis of schizophrenia and autism. *Psychiatric genetics.* 2015; 25(2):79-87.
 49. Karami-Mohajeri S, Ahmadipour A, Rahimi HR, Abdollahi M. Adverse effects of organophosphorus pesticides on the liver: A brief summary of four decades of research. *Archives of Industrial Hygiene and Toxicology.* 2017; 68(4):261-275.
 50. Khazaie S, Jafari M, Heydari J, *et al.* Modulatory effects of vitamin C on biochemical and oxidative changes induced by acute exposure to diazinon in rat various tissues: Prophylactic and therapeutic roles. *Journal of Animal Physiology and Animal Nutrition.* 2019; 103(5):1619-1628.
 51. Tatipamula VB, Kukavica B. Protective effects of extracts of lichen *Dirinaria consimilis* (Stirton) D.D. Awasthi in bifenthrin- and diazinon-induced oxidative stress in rat erythrocytes *in vitro*. *Drug and Chemical Toxicology.* 2020; 45(2):680-687.
 52. Tahmasebi K, Jafari M, Heydari J, *et al.* Tissues toxicity attenuation by vitamin E on oxidative damage induced by diazinon. *Environmental Analysis Health and Toxicology.* 2022; 37(4):e2022036.
 53. Mehta A, Verma RS, Srivastava N. Chlorpyrifos induced alterations in the levels of hydrogen peroxide, nitrate and nitrite in rat brain and liver. *Pesticide Biochemistry and Physiology.* 2009; 94(2-3):55-59.
 54. El-Shenawy NS, El-Salmy F, Al-Eisa RA, El-Ahmary B. Amelioratory effect of vitamin E on organophosphorus insecticide diazinon-induced oxidative stress in mice liver. *Pesticide Biochemistry and Physiology.* 2009; 96(2):101-107.
 55. Hassani S, Maqbool F, Salek-Maghsoudi A, *et al.* Alteration of hepatocellular antioxidant gene expression pattern and biomarkers of oxidative damage in diazinon-induced acute toxicity in Wistar rat: A time-course mechanistic study. *PubMed.* 2018; 17:57-71.
 56. Vahidirad M, Arab-Nozari M, Mohammadi H, Zamani E, Shaki F. Protective effect of captopril against diazinon induced nephrotoxicity and neurotoxicity via inhibition of ROS-NO pathway. *Drug and Chemical Toxicology.* 2017; 41(3):287-293.
 57. Wang W, Luo SM, Ma JY, Shen W, Yin S. Cytotoxicity and DNA Damage Caused from Diazinon Exposure by Inhibiting the PI3K-AKT Pathway in Porcine Ovarian Granulosa Cells. *Journal of Agricultural and Food Chemistry.* 2018; 67(1):19-31.
 58. Nassar WM, El-Kholy WM, El-Sawi MR, *et al.* Ameliorative Effect of Thymoquinone and Thymoquinone Nanoparticles against Diazinon-Induced Hepatic Injury in Rats: A Possible Protection Mechanism. *Toxics.* 2023; 11(9):783.
 59. Naderi N, Souri M, Nasr-Esfahani MH, Hajian M, Nazem MN. *Ferulago angulata* extract alleviates testicular toxicity in male mice exposed to diazinon and lead. *Tissue and Cell.* 2023; 85:102257.
 60. Ali NI, Salem LM, Elateek SY, Khalil WK. Modulation impact of Diazinon forms on gene expression profile and DNA damage pathway in male mice. *Journal of Applied Pharmaceutical Science.* 2020; 10(8):067-074.
 61. Li W, Liu Y, Duan J, Van Leeuwen J, Saint CP. UV and UV/H₂O₂ treatment of diazinon and its influence on disinfection byproduct formation following chlorination. *Chemical Engineering Journal.* 2015; 274:39-49.
 62. Rems L, Viano M, Kasimova MA, Miklavčič D, Tarek M. The contribution of lipid peroxidation to membrane permeability in electroporation: A molecular dynamics study. *Bioelectrochemistry.* 2018; 125:46-57.
 63. Abdel-Daim MM. Synergistic protective role of ceftriaxone and ascorbic acid against subacute diazinon-induced nephrotoxicity in rats. *Cytotechnology.* 2014; 68(2):279-289.
 64. Abdelkhalik NKM, Eissa IAM, Ahmed E, *et al.* Protective role of dietary *Spirulina platensis* against diazinon-induced Oxidative damage in Nile tilapia; *Oreochromis niloticus*. *Environmental Toxicology and Pharmacology.* 2017; 54:99-104.

65. Ivanovic S, Borozan N, Jankovic R, *et al.* Functional and histological changes of the pancreas and the liver in the rats after the acute and subacute administration of diazinon. *Vojnosanitetski Pregled.* 2021; 78(9):955-963.
66. Beydilli H, Yilmaz N, Cetin ES, *et al.* Evaluation of the protective effect of silibinin against diazinon induced hepatotoxicity and Free-Radical damage in rat liver. *Iranian Red Crescent Medical Journal.* 2015; 17(4).
67. Al-Attar AM. Effect of grapeseed oil on diazinon-induced physiological and histopathological alterations in rats. *Saudi Journal of Biological Sciences.* 2015; 22(3):284-292.
68. Lari P, Abnous K, Imenshahidi M, Rashedinia M, Razavi M, Hosseinzadeh H. Evaluation of diazinon-induced hepatotoxicity and protective effects of crocin. *Toxicology and Industrial Health.* 2013; 31(4):367-376.
69. Pourtaji A, Robati RY, Lari P, Hosseinzadeh H, Ramezani M, Abnous K. Proteomics screening of adenosine triphosphate-interacting proteins in the liver of diazinon-treated rats. *Human & Experimental Toxicology.* 2016; 35(10):1084-1092.
70. Kalender S, Ogutcu A, Uzunhisarcikli M, *et al.* Diazinon-induced hepatotoxicity and protective effect of vitamin E on some biochemical indices and ultrastructural changes. *Toxicology.* 2005; 211(3):197-206.
71. Ghasemzadeh L, Mohajereani H, Nasri S, Rostami A. The effect of diazinon exposure on hepatic tissue and enzymes in male frog *Rana sridibunda*. *Progress in Biological Sciences.* 2015; 5(2):223-232.
72. Ferguson MA, Vaidya VS, Bonventre JV. Biomarkers of nephrotoxic acute kidney injury. *Toxicology.* 2008; 245(3):182-193.
73. Biewenga GPh, Haenen GRMM, Bast A. The pharmacology of the antioxidant lipoic acid. *General Pharmacology the Vascular System.* 1997; 29(3):315-331.
74. Ajibade TO, Oyagbemi AA, Omobowale TO, Asenuga ER, Afolabi JM, Adedapo AA. Original article. Mitigation of diazinon-induced cardiovascular and renal dysfunction by gallic acid. *Interdisciplinary Toxicology.* 2016; 9(2):66-77.
75. Shah MD, Iqbal M. Diazinon-induced oxidative stress and renal dysfunction in rats. *Food and Chemical Toxicology.* 2010; 48(12):3345-3353.
76. Al-Attar AM, Zeid IMA. Effect of Tea (*Camellia sinensis*) and Olive (*Olea europaea*L.) Leaves Extracts on Male Mice Exposed to Diazinon. *BioMed Research International.* 2013; 2013:1-6.
77. Delorenzi Schons D, Leite GAA. Malathion or diazinon exposure and male reproductive toxicity: A systematic review of studies performed with rodents. *Critical Reviews in Toxicology.* 2023; 53(8):06-520.
78. Cobilinschi C, Țincu RC, Cobilinschi CO, *et al.* Histopathological features of low-dose organophosphate exposure. *Romanian Journal of Morphology and Embryology.* 2020; 61(2):423-432.
79. Anbarkeh FR, Nikravesh MR, Jalali M, Sadeghnia HR, Sargazi Z, Mohammadzadeh L. Single dose effect of diazinon on biochemical parameters in testis tissue of adult rats and the protective effect of vitamin E. *PubMed.* 2014; 12(11):731-736.
80. Harchegani AB, Rahmani A, Tahmasbpour E, Kabootaraki HB, Rostami H, Shahriary A. Mechanisms of diazinon effects on impaired spermatogenesis and male infertility. *Toxicology and Industrial Health.* 2018; 34(9):653-664.
81. Piña-Guzmán B, Solís-Heredia MJ, Quintanilla-Vega B. Diazinon alters sperm chromatin structure in mice by phosphorylating nuclear protamines. *Toxicology and Applied Pharmacology.* 2005; 202(2):189-198.
82. Salazar-Arredondo E, De Jesús Solís-Heredia M, Rojas-García E, Hernández-Ochoa I, Quintanilla-Vega B. Sperm chromatin alteration and DNA damage by methyl-parathion, chlorpyrifos and diazinon and their oxon metabolites in human spermatozoa. *Reproductive Toxicology.* 2008; 25(4):455-460.
83. Aluigi MG, Guida C, Falugi C. Apoptosis as a specific biomarker of diazinon toxicity in NTERA2-D1 cells. *Chemico-Biological Interactions.* 2010; 187(1-3):299-303.
84. Shiri M, Navaei-Nigjeh M, Baeeri M, *et al.* Blockage of both the extrinsic and intrinsic pathways of diazinon-induced apoptosis in PaTu cells by magnesium oxide and selenium nanoparticles. *International Journal of Nanomedicine.* 2016; 11:6239-6250.
85. Amaral A, Lourenço B, Marques M, Ramalho-Santos J. Mitochondria functionality and sperm quality. *Reproduction.* 2013; 146(5):R163-R174.
86. Piomboni P, Focarelli R, Stendardi A, Ferramosca A, Zara V. The role of mitochondria in energy production for human sperm motility. *Andrology.* 2011; 35(2):109-124.
87. Ebadimanas G, Najafi G. Reduction of Oxidative Stress and Testicular Tissue Damage in Wistar Rat Treated with Diazinon by Quercetin. *Journal of Kermanshah University of Medical Sciences.* 2024; 28(3).
88. Abou Hasan F, Mutlu HS, Özdemir İ, Kotil T. Effects of diazinon on the ovarian tissue of rats: A histochemical and ultrastructural study. *Journal of Molecular Histology.* 2024; 55(6):1-13.
89. Siavashpour A, Ghasemi Y, Khalvati B, Jeivad F, Azarpira N, Niknahad H. Diazinon interrupts ovarian steroidogenic acute regulatory (STAR) gene transcription in Gonadotropin-Stimulated RAT model, 2018. Available at: <https://pmc.ncbi.nlm.nih.gov/articles/PMC5985171>.
90. Jarc E, Petan T. Focus: Organelles: Lipid droplets and the management of cellular stress. *The Yale Journal of Biology and Medicine.* 2019; 92(3):435.
91. Boyda J, Hawkey AB, Holloway ZR, Trevisan R, Di Giulio RT, Levin ED. The organophosphate insecticide diazinon and aging: Neurobehavioral and mitochondrial effects in zebrafish exposed as embryos or during aging. *Neurotoxicology and Teratology.* 2021; 87:107011.
92. Zhao S, Wesseling S, Spenkelink B, Rietjens IMCM. Physiologically based kinetic modelling based prediction of *in vivo* rat and human acetylcholinesterase (AChE) inhibition upon exposure to diazinon. *Archives of Toxicology.* 2021; 95(5):1573-1593.
93. Papandreou MA, Dimakopoulou A, Linardaki ZI, *et al.* Effect of a polyphenol-rich wild blueberry extract on cognitive performance of mice, brain antioxidant markers and acetylcholinesterase activity. *Behavioural Brain Research.* 2009; 198(2):352-358.
94. Baltazar MT, Dinis-Oliveira RJ, De Lourdes Bastos M,

- Tsatsakis AM, Duarte JA, Carvalho F. Pesticides exposure as etiological factors of Parkinson's disease and other neurodegenerative diseases—A mechanistic approach. *Toxicology Letters*. 2014; 230(2):85-103.
95. Ahmed MAE, Ahmed HI, El-Morsy EM. Melatonin protects against Diazinon-Induced neurobehavioral changes in rats. *Neurochemical Research*. 2013; 38(10):2227-2236.
 96. Barrett K, Jaward FM. A review of endosulfan, dichlorvos, diazinon, and diuron – pesticides used in Jamaica. *International Journal of Environmental Health Research*. 2012; 22(6):481-499.
 97. Win-Shwe TT, Nakajima D, Ahmed S, Fujimaki H. Impairment of novel object recognition in adulthood after neonatal exposure to diazinon. *Archives of Toxicology*. 2013; 87(4):753-762.
 98. Slotkin TA, Skavicus S, Ko A, Levin ED, Seidler FJ. Perinatal diazinon exposure compromises the development of acetylcholine and serotonin systems. *Toxicology*. 2019; 424:152240.
 99. Hawkey A, Pippen E, White H, *et al.* Gestational and perinatal exposure to diazinon causes long-lasting neurobehavioral consequences in the rat. *Toxicology*. 2020; 429:152327.
 100. Vatanparast J, Naseh M, Baniasadi M, Haghdoost-Yazdi H. Developmental exposure to chlorpyrifos and diazinon differentially affect passive avoidance performance and nitric oxide synthase-containing neurons in the basolateral complex of the amygdala. *Brain Research*. 2013; 1494:17-27.
 101. Wu N, Wang F, Jin Z, *et al.* Effects of GABAB receptors in the insula on recognition memory observed with intellicage. *Behavioral and Brain Functions*. 2017; 13(1).
 102. Karimani A, Ramezani N, Goli AA, Shirazi MHN, Nourani H, Jafari AM. Subchronic neurotoxicity of diazinon in albino mice: Impact of oxidative stress, AChE activity, and gene expression disturbances in the cerebral cortex and hippocampus on mood, spatial learning, and memory function. *Toxicology Reports*. 2021; 8:1280-1288.
 103. Afshari S, Sarailoo M, Asghariazar V, Safarzadeh E, Dadkhah M. Persistent diazinon induced neurotoxicity: The effect on inhibitory avoidance memory performance, amyloid precursor proteins, and TNF- α levels in the prefrontal cortex of rats. *Human & Experimental Toxicology*. 2024; 43.
 104. Pizzurro DM, Dao K, Costa LG. Diazinon and diazoxon impair the ability of astrocytes to foster neurite outgrowth in primary hippocampal neurons. *Toxicology and Applied Pharmacology*. 2014; 274(3):372-382.
 105. Razavi BM, Hosseinzadeh H, Movassaghi AR, Imenshahidi M, Abnous K. Protective effect of crocin on diazinon induced cardiotoxicity in rats in subchronic exposure. *Chemico-Biological Interactions*. 2013; 203(3):547-555.
 106. Griendling KK, Sorescu D, Ushio-Fukai M. NAD(P)H oxidase. *Circulation Research*. 2000; 86(5):494-501.
 107. McKinlay R, Plant JA, Bell JNB, Voulvoulis N. Endocrine disrupting pesticides: Implications for risk assessment. *Environment International*. 2008; 34(2):168-183.
 108. Rattan S, Zhou C, Chiang C, Mahalingam S, Brehm E, Flaws JA. Exposure to endocrine disruptors during adulthood: Consequences for female fertility. *Journal of Endocrinology*. 2017; 233(3):R109-R129.
 109. Senthilkumaran B. Pesticide- and sex steroid analogue-induced endocrine disruption differentially targets hypothalamo-hypophyseal-gonadal system during gametogenesis in teleosts – A review. *General and Comparative Endocrinology*. 2015; 219:136-142.
 110. Ghajari G, Moosavi R. Evaluation of the effects of diazinon toxin on some reproductive parameters in male rats. *Personalized Medicine Journal*. 2022; 7(25):30-35.
 111. Pournourmohammadi S, Farzami B, Ostad SN, Azizi E, Abdollahi M. Effects of malathion subchronic exposure on rat skeletal muscle glucose metabolism. *Environmental toxicology and Pharmacology*. 2005; 19(1):191-196.
 112. Čolović MB, Vasić VM, Avramović NS, Gajić MM, Djurić DM, Krstić DZ. *In vitro* evaluation of neurotoxicity potential and oxidative stress responses of diazinon and its degradation products in rat brain synaptosomes. *Toxicology Letters*. 2015; 233(1):29-37.
 113. Layali I, Tahmasbpour E, Joulai M, Jorsaraei SGA, Farzanegi P. Total antioxidant capacity and lipid peroxidation in semen of patient with hyperviscosity. *DOAJ DOAJ: Directory of Open Access Journals*. 2015; 16(4):554-559.
 114. Moallem SA, Hariri AT, Mahmoudi M, Hosseinzadeh H. Effect of aqueous extract of *Crocus sativus* L. (saffron) stigma against subacute effect of diazinon on specific biomarkers in rats. *Toxicology and Industrial Health*. 2014; 30(2):141-146.
 115. Danaei GH, Karami M. Protective effect of thymoquinone against diazinon-induced hematotoxicity, genotoxicity and immunotoxicity in rats. *Environmental Toxicology and Pharmacology*. 2017; 55:217-222.
 116. Abdel-Diam MM, Samak DH, El-Sayed YS, Aleya L, Alarifi S, Alkahtani S. Curcumin and quercetin synergistically attenuate subacute diazinon-induced inflammation and oxidative neurohepatic damage, and acetylcholinesterase inhibition in albino rats. *Environmental Science and Pollution Research*. 2019; 26(4):3659-3665.
 117. Hariri AT, Moallem SA, Mahmoudi M, Memar B, Hosseinzadeh H. Sub-acute effects of diazinon on biochemical indices and specific biomarkers in rats: Protective effects of crocin and safranal. *Food and Chemical Toxicology*. 2010; 48(10):2803-2808.
 118. Abdel-Daim MM, Abushouk AI, Alkhalf MI, *et al.* Antagonistic effects of *Spirulina platensis* on diazinon-induced hemato-biochemical alterations and oxidative stress in rats. *Environmental Science and Pollution Research*. 2018; 25(27):27463-27470.
 119. Alluwaimi AM, Hussein Y. Diazinon immunotoxicity in mice: Modulation of cytokines level and their gene expression. *Toxicology*. 2007; 236(1-2):123-131.
 120. Neishabouri EZ, Hassan ZM, Azizi E, Ostad SN. Evaluation of immunotoxicity induced by diazinon in C57bl/6 mice. *Toxicology*. 2004; 196(3):173-179.
 121. Díaz-Resendiz KJG, Ortiz-Lazareno PC, Covantes-Rosales CE, *et al.* Effect of diazinon, an organophosphate pesticide, on signal transduction and death induction in mononuclear cells of Nile tilapia fish (*Oreochromis niloticus*). *Fish & Shellfish Immunology*. 2019; 89:12-17.
 122. Girón-Pérez MI, Santerre A, Gonzalez-Jaime F, *et al.*

- Immunotoxicity and hepatic function evaluation in Nile tilapia (*Oreochromis niloticus*) exposed to diazinon. *Fish & Shellfish Immunology*. 2007; 23(4):760-769.
123. Khoshbavar-Rostami HA, Soltani M, Hassan HMD. Immune response of great sturgeon (*Huso huso*) subjected to long-term exposure to sublethal concentration of the organophosphate, diazinon. *Aquaculture*. 2006; 256(1-4):88-94.
124. Ruan H, Lodish HF. Insulin resistance in adipose tissue: Direct and indirect effects of tumor necrosis factor- α . *Cytokine & Growth Factor Reviews*. 2003; 14(5):447-455.
125. Reinehr T, Andler W. Cortisol and Its Relation to Insulin Resistance before and after Weight Loss in Obese Children. *Hormone Research in Paediatrics*. 2004; 62(3):107-112.
126. Ueyama J, Kamijima M, Asai K, *et al.* Effect of the organophosphorus pesticide diazinon on glucose tolerance in type 2 diabetic rats. *Toxicology Letters*. 2008; 182(1-3):42-47.
127. Abdollahi M, Moridani MY, Aruoma OI, Mostafalou S. Oxidative stress in aging. *Oxidative Medicine and Cellular Longevity*. 2014; 2014:1-2.
128. Farkhondeh T, Aschner M, Sadeghi M, *et al.* The effect of diazinon on blood glucose homeostasis: A systematic and meta-analysis study. *Environmental Science and Pollution Research*. 2021; 28(4):4007-4018.